

In Vitro Activity of Tigecycline Against In-patient and Out-patient Enterobacteriaceae Collected From Australia - T.E.S.T. Program 2006

S. Bouchillon¹, B. Johnson¹, R. Badal¹, M. Hackel¹, J. Johnson¹, D. Hoban¹, M. Dowzicky²

¹International Health Management Associates, Schaumburg, IL, USA

²Wyeth Pharmaceuticals, Collegeville, PA, USA

IHMA, Inc.
2122 Palmer Dr.
Schaumburg, IL 60173
Tel: (847) 303-5003
Fax: (847) 303-5601
www.ihmainc.com

REVISED ABSTRACT

Background: Tigecycline, a member of a new class of antimicrobials (glycylcyclines), has been shown to have potent broad spectrum activity against community and hospital acquired infections. The T.E.S.T. program determined the in vitro activity of tigecycline compared to most commonly prescribed broad spectrum antimicrobials against gram-negative and gram-positive species collected from 3 investigative sites in Australia from 2005 to 2006. **Methods:** A total of 241 clinical *Enterobacteriaceae* isolates from Australia were identified to the species level and confirmed by the central laboratory. Minimum Inhibitory Concentration (MICs) were determined by each site using broth microdilution panels and interpreted according to CLSI guidelines. **Results:** Cumulative data results are in the table as follows, with individual species data to be presented:

	<i>Enterobacteriaceae</i>			
	In patients (n=173)		Out patients (n=68)	
	%S	MIC ₉₀	%S	MIC ₉₀
Tigecycline	96.5	1	100	1
Amikacin	100	4	100	4
Cefepime	98.3	2	100	≤0.5
Ceftazidime	80.9	32	95.6	≤8
Imipenem	100	1	100	1
Levofloxacin	98.3	0.25	100	0.12
Minocycline	90.2	4	94.1	4
PipTazo	84.4	64	92.6	4

[†]Tigecycline susceptibility ≤2 mcg/mL (Tygacil[®], FDA package insert, 2005)

Conclusion: Tigecycline's in vitro activity was comparable to or greater than most commonly prescribed broad spectrum antimicrobials without any categorical susceptibility change between In- and Out-patient bacterial study strains. Tigecycline in vitro activity was comparable to most of the study broad spectrum antimicrobials. The presented data suggest that tigecycline may be an effective and reliable therapeutic option against nosocomial or community enteric pathogens.

INTRODUCTION

Tigecycline is a novel antimicrobial with expanded broad-spectrum activity from a new class of compounds, the glycylcyclines. Tigecycline inhibits protein synthesis by binding to the 30S ribosomal subunit. Although it is perceived to be bacteriostatic, its anti-bacterial activity is significant and has shown some bactericidal activity against key targeted pathogens [1,2]. Tigecycline was developed to provide activity against tetracycline and multi-drug-resistant gram-positive pathogens and has demonstrated significant broad-spectrum activity against aerobic and anaerobic gram-positive and gram-negative microorganisms [2-4].

Tigecycline resistance is very infrequent and is also difficult to induce in the laboratory [5, 6] with a selection frequency observed at less than 10⁻⁹ [3, 5, 7]. With the exception of *P. aeruginosa*, tetracycline-resistant bacteria with either tetracycline efflux pumps or ribosomal protective features are sensitive to tigecycline [2-4, 7-11]. Tigecycline has shown to be highly effective against multi-resistant *Acinetobacter* spp., particularly *A. baumannii* that are commonly associated with serious nosocomial infections. Similar activity has been observed against *Enterobacteriaceae*, even extended-spectrum beta-lactamase (ESBL) and AmpC producing strains [10]. Tigecycline has demonstrated MIC₉₀ values of <0.5 mcg/ml against methicillin-resistant *Staphylococcus aureus* (MRSA) and other gram-positive organisms [2, 4-6]. Tigecycline has shown potent activity against animal models infected with selected strains of multi-drug resistant *Enterococcus faecium* and *Enterococcus faecalis* [4, 5] with diverse genotypes van-A, -B and -C [6].

This study was designed to communicate clinical laboratory in vitro experience with activity of tigecycline in a limited number against isolates collected from three medical centers in Australia.

MATERIALS & METHODS

- All isolates were derived from blood, respiratory tract, urine (no more than 25% of all isolates), skin, wound, fluids and few other defined sources. Only one isolate per patient was accepted. Isolates were identified to genus and species by the local laboratory. Each site tested the isolates using broth microdilution.
- 500 clinical isolates were collected tested between January 2005 - December 2006 from three study centers from Australia. Of these, 241 had designations of In-Patient and Out-patient locations.
- Custom broth microdilution panels were supplied by MicroScan (Dade MicroScan, Sacramento, CA, USA) with the following antimicrobial agents and concentrations (expressed in mcg/mL): amoxicillin/clavulanic acid (0.03-32); piperacillin/tazobactam (0.06-128); levofloxacin (0.008-32); ceftriaxone (0.03-64); cefepime (0.5-32); ampicillin (0.06-32); amikacin (0.5-64); minocycline (0.25-16); ceftazidime (8-32); tigecycline (0.008-16); and imipenem (0.06-16).
- MIC interpretive criteria followed published guidelines established by the CLSI where applicable [12]. MIC interpretive criteria for Tigecycline followed published guidelines established by the FDA where applicable [13].
- Quality control of broth microdilution panels followed manufacturer's and CLSI guidelines using the following ATCC strains: *E. coli* ATCC 25922; *K. pneumoniae* ATCC 700603; and *Pseudomonas aeruginosa* ATCC 27853.
- The collection and transportation of organisms and the confirmation of identification, as well as, construction and management of a centralized database were conducted and coordinated by Laboratories International for Microbiology Studies (LIMS), a subsidiary of International Health Management Associates, Inc. (IHMA, Schaumburg, IL, USA).

REFERENCES

- 1 Sum, P.E. and P. Petersen, Synthesis and structure-activity relationship of novel glycylcycline derivatives leading to the discovery of GAR-936. *Bioorg Med Chem Lett*, 1999, 9(10): p. 1459-62.
- 2 Abbanat, D., M. Macielag, and K. Bush, Novel antibacterial agents for the treatment of serious Gram-positive infections. *Expert Opin Investig Drugs*, 2003, 12(3): p. 379-99.
- 3 Betriu, C., et al., In vitro activities of tigecycline (GAR-936) against recently isolated clinical bacteria in Spain. *Antimicrob Agents Chemother*, 2002, 46(3): p. 892-5.
- 4 Gates, A.C. and R.N. Jones, Antimicrobial activity and spectrum of the new glycylcycline, GAR-936 tested against 1,203 recent clinical bacterial isolates. *Diagn Microbiol Infect Dis*, 2000, 36(1): p. 19-36.
- 5 Henwood, C.J., et al., Antibiotic resistance among clinical isolates of *Acinetobacter* in the UK, and in vitro evaluation of tigecycline (GAR-936). *J Antimicrob Chemother*, 2002, 49(3): p. 479-87.
- 6 Chopra, I., New developments in tetracycline antibiotics: glycylcyclines and tetracycline efflux pump inhibitors. *Drug Resist Updat*, 2002, 5(3-4): p. 119-25.
- 7 Projan, S.J., Preclinical pharmacology of GAR-936, a novel glycylcycline antibacterial agent. *Pharmacotherapy*, 2000, 20(9 Pt 2): p. 219S-223S; discussion 224S-228S.
- 8 Biedenbach, D.J., M.L. Beach, and R.N. Jones, In vitro antimicrobial activity of GAR-936 tested against antibiotic-resistant gram-positive blood stream infection isolates and strains producing extended-spectrum beta-lactamases. *Diagn Microbiol Infect Dis*, 2001, 40(4): p. 173-7.
- 9 Patel, R., et al., In vitro activity of GAR-936 against vancomycin-resistant enterococci, methicillin-resistant *Staphylococcus aureus* and penicillin-resistant *Streptococcus pneumoniae*. *Diagn Microbiol Infect Dis*, 2000, 38(3): p. 177-9.
- 10 Petersen, P.J., et al., In vitro and in vivo antibacterial activities of a novel glycylcycline, the 9-t-butylglycylamido derivative of minocycline (GAR-936). *Antimicrob Agents Chemother*, 1999, 43(4): p. 738-44.
- 11 Petersen, P.J., et al., In vitro and in vivo activities of tigecycline (GAR-936), daptomycin, and comparative antimicrobial agents against glycopeptide-intermediate *Staphylococcus aureus* and other resistant gram-positive pathogens. *Antimicrob Agents Chemother*, 2002, 46(8): p. 2595-601.
- 12 Clinical Laboratory Standards Institute (CLSI), Performance Standards for Antimicrobial Susceptibility Testing; 16th Informational Supplement. CLSI document M100-S16. Wayne, PA, 2006.
- 13 Tygacil[®], 2005. Tigecycline FDA package insert.

ACKNOWLEDGEMENTS

We are grateful to the laboratory staffs at Flinders Medical Centre, The Prince Charles Hospital Campus and Westmead Hospital for their data contributions to this study. Special thanks to Brian Johnson of IHMA for the technical reproduction of this publication. This study was supported by a grant from Wyeth Pharmaceuticals.

RESULTS

The results of this study are presented in the following tables:

Table 1. Gram-negative antibiogram for tigecycline and 10 comparators with MIC₉₀ (mcg/mL) and percent susceptible (%) against 251 selected *Enterobacteriaceae*.

Organisms	N=	MIC ₉₀ in mcg/mL (% Susceptible) [*]						
		Tige	Amik	Cefep	Ceftaz	Imip	Levo	PipTazo
<i>Enterobacteriaceae</i> [†]	251	1(97.6)	4(100)	2(98.4)	32(84.9)	1(100)	0.25(98.8)	64(86.5)
<i>E. aerogenes</i>	26	0.5(96.2)	2(100)	≤0.5(100)	32(65.4)	1(100)	0.12(100)	32(88.5)
<i>E. cloacae</i>	48	1(95.8)	4(100)	8(93.8)	>32(52.1)	1(100)	0.5(97.9)	64(58.3)
<i>E. gergoviae</i>	1	0.5(100)	1(100)	8(100)	>32(0)	0.5(100)	0.12(100)	128(0)
<i>E. coli</i>	75	0.25(100)	8(100)	≤0.5(100)	≤8(98.7)	0.5(100)	0.06(98.7)	2(97.3)
<i>K. oxytoca</i>	29	0.5(100)	2(100)	2(100)	≤8(96.6)	0.5(100)	0.12(100)	>128(75.9)
<i>K. pneumoniae</i>	42	1(97.6)	2(100)	≤0.5(97.6)	≤8(95.2)	0.5(100)	0.25(97.6)	4(97.6)
<i>K. pneumoniae</i> , ESBL	1	1(100)	4(100)	>32(0)	16(0)	0.25(100)	>8(0)	8(100)
<i>S. marcescens</i>	29	2(96.6)	8(100)	≤0.5(100)	≤8(96.6)	2(100)	0.5(100)	4(100)
<i>S. odorifera</i>	1	4(0)	2(100)	≤0.5(100)	≤8(100)	0.5(100)	0.12(100)	2(100)

^{*} Susceptibility defined by CLSI document M100-S16 (2006), where available.

[†] All *Enterobacter*, *Escherichia*, *Klebsiella* and *Serratia* combined.

Table 2. In vitro activity of tigecycline and comparators against selected *Enterobacteriaceae* for in-patient vs out-patient populations^{*}

Organism	Drug	In-Patient					Out-Patient					p-Value [†]
		%Sus	%Res	Mean MIC	MIC ₉₀	%Sus	%Res	Mean MIC	MIC ₉₀			
<i>Enterobacter aerogenes</i> (n=IP 20; n=OP 5)	Tigecycline	95	0	0.7	2	100	0	0.4	0.5	0.4501		
	Amikacin	100	0	1.65	2	100	0	1.6	2	0.89		
	Cefepime	100	0	0.549	1	100	0	0.499	≤0.5	0.4819		
	Ceftazidime	60	15	13.6	32	80	20	12.799	32	0.8611		
	Imipenem	100	0	0.687	1	100	0	0.8	1	0.457		
	Levofloxacin	100	0	0.054	0.06	100	0	0.066	0.12	0.6402		
	PipTazo	90	0	9.25	16	80	0	14	64	0.5193		
	<i>Enterobacter cloacae</i> (n=IP 35; n=OP 11)	Tigecycline	95	4.5	1.021	1	100	0	0.681	1	0.536	
Amikacin		100	0	1.857	2	100	0	1.727	2	0.7613		
Cefepime		94.3	5.7	3.185	4	100	0	1.363	2	0.4251		
Ceftazidime		45.7	42.9	19.2	>32	81.8	18.2	12.366	32	0.0824		
Imipenem		100	0	0.55	1	100	0	0.727	1	0.247		
Levofloxacin		97.1	2.9	0.328	0.5	100	0	0.169	0.5	0.7011		
PipTazo		54.3	8.6	32.228	64	81.8	9.1	16.136	32	0.2356		
<i>Escherichia coli</i> (n=IP 43; n=OP 29)		Tigecycline	100	0	0.174	0.25	100	0	0.153	0.25	0.2516	
	Amikacin	100	0	3.023	4	100	0	3.034	8	0.9811		
	Cefepime	100	0	0.581	≤0.5	100	0	0.499	≤0.5	0.4154		
	Ceftazidime	97.7	2.3	8.558	≤8	100	0	7.999	≤8	0.4154		
	Imipenem	100	0	0.279	0.5	100	0	0.301	0.5	0.4207		
	Levofloxacin	97.7	2.3	0.253	0.25	100	0	0.035	0.06	0.3396		
	PipTazo	95.3	4.7	7.331	4	100	0	1.198	2	0.2267		
	<i>Klebsiella oxytoca</i> (n=IP 20; n=OP 9)	Tigecycline	100	0	0.412	0.5	100	0	0.277	0.5	0.3484	
Amikacin		100	0	1.65	2	100	0	1.666	2	0.9519		
Cefepime		100	0	0.949	2	100	0	0.888	4	0.9228		
Ceftazidime		95	0	8.399	≤8	100	0	7.999	≤8	0.5123		
Imipenem		100	0	0.387	0.5	100	0	0.333	1	0.5248		
Levofloxacin		100	0	0.15	0.25	100	0	0.03	0.03	0.4235		
PipTazo		75	25	33.02	>128	77.8	22.2	29.278	>128	0.8693		
<i>Klebsiella pneumoniae</i> (n=IP 30; n=OP 10)		Tigecycline	96.7	0	0.783	1	100	0	0.575	1	0.3986	
	Amikacin	100	0	1.333	2	100	0	1.2	2	0.62		
	Cefepime	96.7	3.3	1.549	≤0.5	100	0	0.499	≤0.5	0.5705		
	Ceftazidime	93.3	0	8.533	≤8	100	0	7.999	≤8	0.4152		
	Imipenem	100	0	0.325	0.5	100	0	0.3	0.5	0.5517		
	Levofloxacin	96.7	3.3	0.406	0.25	100	0	0.092	0.25	0.5088		
	PipTazo	96.7	0	4.066	4	100	0	2.3	2	0.6332		
	<i>Serratia marcescens</i> (n=IP 23; n=OP 4)	Tigecycline	95.7	0	1.369	2	100	0	1	1	0.3364	
Amikacin		100	0	4.347	8	100	0	1.5	2	0.1677		
Cefepime		100	0	0.847	≤0.5	100	0	0.499	≤0.5	0.6651		
Ceftazidime		95.7	4.3	9.043	≤8	100	0	7.999	≤8	0.6851		
Imipenem		100	0	0.875	2	100	0	0.666	1	0.5229		
Levofloxacin		100	0	0.192	0.5	100	0	0.06	0.12	0.1042		
PipTazo		100	0	2.913	4	100	0	1.25	2	0.341		

^{*} Susceptibilities defined by CLSI document M100-S16 (2006), where available. Tigecycline breakpoints defined in FDA package insert (Tygacil[®], 2005).

[†] One-way ANOVA.

CONCLUSIONS

- Tigecycline inhibited 97.6% of all *Enterobacteriaceae* tested in vitro at a MIC₉₀ of 1 mcg/mL. Tigecycline's MIC₉₀ of 1 mcg/mL was equivalent to imipenem and 2- to 64-fold better than the beta-lactams and beta-lactam/beta-lactamase inhibitor combinations. Only levofloxacin had a lower MIC₉₀ at 0.25 mcg/mL against all *Enterobacteriaceae* tested.
- Tigecycline activity exhibited small, but statistically significant (p-value <0.5), increases in mean MIC values for in-patient compared to out-patient *Enterobacteriaceae* study species. Similar increases in mean MIC values were seen in two or more study species for cefepime, ceftazidime, imipenem, levofloxacin and piperacillin-tazobactam. Tigecycline inhibited between 95% and 100% of all study species at or below its susceptibility breakpoint of 2 mcg/mL regardless of the source location, in-patient or out-patient, of the causative pathogen.
- The in vitro activity of tigecycline in this study suggests that tigecycline is an effective agent with excellent antimicrobial activity in this single clinical laboratory against common gram-negative nosocomial and community acquired pathogens.